

FORM PTO-1390

U.S. DEPARTMENT OF COMMERCE PATENT AND TRADEMARK OFFICE
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 DESIGNATED/ELECTED OFFICE (DO/EO/US)
 CONCERNING A FILING UNDER 35 U.S.C. 371

ATTORNEY'S DOCKET NUMBER:
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INTERNATIONAL APPLICATION NO.
 PCT/NZ00/00001

INTERNATIONAL FILING DATE:
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PRIORITY DATE CLAIMED:

TITLE OF INVENTION: METHOD FOR RECOVERING PIGMENTS FROM ALGAL CULTURES

APPLICANT(S) FOR DO/EO/US: Peter James KEATING

Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information.

1. ☒ This is a **FIRST** submission of items concerning a filing under 35 U.S.C. 371.
 2. ☐ This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C. 371.
 3. ☒ This express request to begin national examination procedures (35 U.S.C. 371(f)) at any time rather than delay examination until the expiration of the applicable time limit set in 35 U.S.C. 371(b) and PCT Articles 22 and 39(1).
 4. ☐ A proper Demand for International Preliminary Examination was made by the 19th month from the earliest claimed priority date.
 5. ☒ A copy of the International Application as filed (35 U.S.C. 371(c)(2))
 - a. ☒ is transmitted herewith (required only if not transmitted by the International Bureau).
 - b. ☐ has been transmitted by the International Bureau. (see attached copy of PCT/IB/308)
 - c. ☐ is not required, as the application was filed in the United States Receiving Office (RO/US).
 6. ☐ A translation of the International Application into English (35 U.S.C. 371(c)(2)).
 7. ☐ Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371(c)(3)).
 - a. ☐ are transmitted herewith (required only if not transmitted by the International Bureau).
 - b. ☐ have been transmitted by the International Bureau.
 - c. ☐ have not been made; however, the time limit for making such amendments has NOT expired.
 - d. ☐ have not been made and will not be made.
 8. ☐ A translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).
 9. ☒ An oath or declaration of the inventor(s) (35 U.S.C. 371(c)(4)).
 10. ☐ A translation of the annexes of the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(c)(5)).
- Item 11. to 16. below concern document(s) or information included:
11. ☐ An Information Disclosure Statement under 37 CFR 1.97 and 1.98.
 12. ☒ An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
 13. ☒ A **FIRST** preliminary amendment.
 14. ☐ A **SECOND** or **SUBSEQUENT** preliminary amendment.
 15. ☐ A substitute specification.
 16. ☐ A change of power of attorney and/or address letter.
 16. ☒ Other items or information: INTERNATIONAL SEARCH REPORT (PCT/ISA/210), APPLICATION DATA SHEET, ABSTRACT

U.S. APPLICATION NO. (if known) 37 CFR 1.401

09/936213

INTERNATIONAL APPLICATION NO.
PCT/NZ00/00001ATTORNEY'S DOCKET NO.
T/JP/00U246US17. ☒ The following fees are submitted:**BASIC NATIONAL FEE (37 CFR 1.492(a)(1)-(5)):**

Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO and International Search Report not prepared by the EPO or JPO \$ 1,000.00

International preliminary examination fee (37 CFR 1.482) not paid to USPTO but International Search Report prepared by the EPO or JPO \$ 860.00

International preliminary examination fee (37 CFR 1.482) not paid to USPTO but international search fee (37 CFR 1.445(a)(2)) paid to USPTO \$ 710.00

International preliminary examination fee (37 CFR 1.482) paid to USPTO but all claims did not satisfy provisions of PCT Article 33(1)-(4) \$ 690.00

International preliminary examination fee (37 CFR 1.482) paid to USPTO and all claims satisfied provisions of PCT Article 33(1)-(4) \$ 100.00

ENTER APPROPRIATE BASIC FEE AMOUNT =

\$ 1000.00

Surcharge of \$130.00 for furnishing the oath or declaration later than months from the earliest claimed priority date (37 CFR 1.492(e)).

\$

CLAIMS

NUMBER FILED

NUMBER EXTRA

RATE

\$

Total claims 14 - 20 = 0 X \$18.00 \$

Independent claims 1 - 3 = 0 X \$80.00 \$

MULTIPLE DEPENDENT CLAIMS(S) (if applicable) + \$270.00 \$

TOTAL OF ABOVE CALCULATIONS =

\$ 1000.00

Reduction of 1/2 for filing by small entity, if applicable. Applicant claims Small Entity Status under 37 CFR 1.27. +

\$

500.00

SUBTOTAL =

\$

500.00

Processing fee of \$130 for furnishing the English translation later than months from the earliest claimed priority date (37 CFR 1.49(f)).

\$

TOTAL NATIONAL FEE =

\$

500.00

Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31). \$40.00 per property +

\$

40.00

TOTAL FEES ENCLOSED =

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540.00

Amount to be
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- a. ☒ A check in the amount of \$ **540.00** to cover the above fees is enclosed.
- b. ☐ Please charge my Deposit Account No. **25-0120** in the amount of \$ to cover the above fees. A duplicate copy of this sheet is enclosed.
- c. ☒ The Commissioner is hereby authorized to charge any additional fees which may be required by 37 CFR 1.16 and 1.17, or credit any overpayment to Deposit Account No. **25-0120**. A duplicate copy of this sheet is enclosed.

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September 10, 2001

By

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PATENTS

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re application of

Peter James KEATING

Box Non-fee Amendment

Serial No. (unknown)

GROUP

Filed herewith

Examiner

METHOD FOR RECOVERING PIGMENTS FROM ALGAL CULTURES

PRELIMINARY AMENDMENT

Commissioner for Patents

Washington, D.C. 20231

Sir:

Prior to the first Official Action and calculation of the filing fee, please amend the above-identified application as follows:

IN THE CLAIMS:

Please amend claims 3-5, 7-8, 10 and 13 as follows:

--3. (Amended) A method as claimed in claim 1 wherein the fat-soluble compounds attach to the metallic iron ore particles via means of adsorption or absorption.

4. (Amended) A method as claimed in claim 1 wherein the wash solution is contacted with the crystalline-metallic-ore-fat-soluble-compound complex by applying the wash solution to the fluidised bed of crystalline metallic ore particles substantially near the bottom of the fluidised bed and at a rate sufficient to maintain the bed in a fluidised state and

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Peter James KEATING

the resultant wash solution containing the fat-soluble compound is collected from near the top of, or above, the fluidised bed of crystalline metallic ore particles.

5.(Amended) A method as claimed in claim 1 further including the step of collecting the crystalline-metallic-ore-fat-soluble-compound complex prior to providing a wash solution and contacting the wash solution with the crystalline-metallic-ore-fat-soluble-compound complex.

7.(Amended) A method as claimed in claim 5 wherein the crystalline-metallic-ore-fat-soluble-compound complex is dried and stored for a period prior to being contacted with the wash solution.

8.(Amended) A method as claimed in claim 1 wherein the fat-soluble compound is present in the aqueous solution within a number of cells and the aqueous solution is a culture media.

10.(Amended) A method as claimed in claim 1 wherein the fat-soluble compound is a natural pigment.

13.(Amended) A substantially pure fat-soluble compound obtained using the method of claim 1.--

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REMARKS

Claims 3-5, 7-8, 10 and 13 were amended to correct multiple dependency. Attached hereto is a marked-up version of the changes made to the claims by the current amendment. The attached page is captioned "VERSION WITH MARKINGS TO SHOW CHANGES MADE".

Respectfully submitted,

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"VERSION WITH MARKINGS TO SHOW CHANGES MADE"

Claims 3-5, 7-8, 10 and 13 have been amended as follows:

3. (Amended) A method as claimed in ~~any one of claims 1 or 2~~ claim 1 wherein the fat-soluble compounds attach to the metallic iron ore particles via means of adsorption or absorption.

4. (Amended) A method as claimed in ~~any one of claims 1 to 3~~ claim 1 wherein the wash solution is contacted with the crystalline-metallic-ore-fat-soluble-compound complex by applying the wash solution to the fluidised bed of crystalline metallic ore particles substantially near the bottom of the fluidised bed and at a rate sufficient to maintain the bed in a fluidised state and the resultant wash solution containing the fat-soluble compound is collected from near the top of, or above, the fluidised bed of crystalline metallic ore particles.

5. (Amended) A method as claimed in ~~any one of claims 1 to 3~~ claim 1 further including the step of collecting the crystalline-metallic-ore-fat-soluble-compound complex prior to providing a wash solution and contacting the wash solution with the crystalline-metallic-ore-fat-soluble-compound complex.

7.(Amended) A method as claimed in ~~any one of claims 5 to 6~~claim 5 wherein the crystalline-metallic-ore-fat-soluble-compound complex is dried and stored for a period prior to being contacted with the wash solution.

8.(Amended) A method as claimed in claim ~~any one of claims 1 to 7~~1 wherein the fat-soluble compound is present in the aqueous solution within a number of cells and the aqueous solution is a culture media.

10.(Amended) A method as claimed in ~~any one of claims 1 to 9~~claim 1 wherein the fat-soluble compound is a natural pigment.

13.(Amended) A substantially pure fat-soluble compound obtained using the method of ~~any one of claims 1 to 12~~.claim 1.

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- 1 -

A METHOD FOR RECOVERING PIGMENTS FROM ALGAL CULTURES

Field

This invention relates to a method of recovering fat soluble compounds, including but not restricted to pigments such as beta-carotene, from solutions, including but not restricted to those solutions containing microalgal cells.

Background

Intensive cultivation of microalgal cells is widely used as the source of a range of biological materials produced by algae including lipids, pigments and protein. A major limitation to the commercial feasibility of manufacturing such materials using algal biotechnology is the fact that microalgal cells exist at relatively low concentrations in water, are of very small size and can be mechanically and osmotically fragile. The harvesting of algal cells and their products at a commercial scale requires processes which concentrate the small algal cells and their constituent chemical products in an efficient manner which is simple, reliable and requires minimal energy inputs.

To date, methods which have been developed involve using either energy requiring processes such as centrifugation and drying, or use low energy processes such as flocculation, settling or algal behavioural responses which are unreliable and inefficient. Other methods require the disintegration of the algal cells which can render any cellular components useless; for example, degradation of valuable components, such as the carotenoids, via oxidation can occur.

One example of a method which may be used to obtain certain cellular components of algal cells, without any adverse degradation of those cellular components, is described in the patent specification relating to PCT/AU82/00165 entitled "Method for Harvesting Algae". This specification concentrates on methods for harvesting and concentrating algae, including *Dunaliella*, from suspensions of a certain salinity, whereby the whole algal cells are adsorbed

- 2 -

onto an appropriate adsorbent media. The principle finding relating to this invention is that algal cell membranes become hydrophobic at salt concentrations above 3M enabling them to adsorb onto substances having a hydrophobic surface. A number of suitable hydrophobic adsorbents are described in this specification. In addition, a process of rendering certain
5 adsorbents hydrophobic, or more hydrophobic, by treatment with silanes for example is described.

In PCT/AU82/00165 the whole-cell-adsorbent-media complex is then processed using organic solvents which damage the cell membrane and potentially which allow cellular components, such as beta-carotene, to be released while the cellular debris and insoluble cell components
10 remain adsorbed to the adsorbent media. The beta-carotene released into the organic solvent in this invention may contains contaminants such as triterpenoids and other lipids and thus further processing is required to isolate only the beta-carotene.

Object

It is an object of the present invention to provide an improved method of extracting fat-soluble
15 compounds or at least to provide the public with a useful choice.

Statement of Invention

In one aspect of the present invention there is provided a method of extracting fat-soluble compounds from aqueous solutions including the steps:

providing an aqueous solution in which a fat-soluble compound is present;

20 providing a bed of crystalline metallic ore particles held in an appropriate vessel;

applying the aqueous solution to the bed of crystalline metallic ore particles substantially near the bottom of the bed at a rate sufficient to form and maintain a fluidised bed of crystalline metallic ore particles;

allowing the fat-soluble compound to attach to the crystalline metallic ore particles to form a
25 crystalline-metallic-ore-fat-soluble-compound complex;

providing a wash solution;

contacting the wash solution with the crystalline-metallic-ore-fat-soluble-compound complex to desorb the fat-soluble compound from the complex;

- 3 -

collecting the wash solution containing the fat-soluble compound; and
isolating the fat-soluble compound from the wash solution.

Preferably the crystalline metallic ore particles are magnetite particles.

- 5 Preferably the wash solution is contacted with the crystalline-metallic-ore-fat-soluble-compound complex by applying the wash solution to the fluidised bed of crystalline metallic ore particles substantially near the bottom of the fluidised bed and at a rate sufficient to maintain the bed in a fluidised state and the resultant wash solution containing the fat-soluble compound is collected from near the top of, or above, the fluidised bed of crystalline metallic ore particles.

- 10 Preferably the method further includes the step of collecting the crystalline-metallic-ore-fat-soluble-compound complex prior to providing a wash solution and contacting the wash solution with the crystalline-metallic-ore-fat-soluble-compound complex.

Preferably the crystalline-metallic-ore-fat-soluble-compound complex is collected from a region substantially near the top of the fluidised bed of crystalline metallic ore particles by means of continuous decantation.

- 15 Preferably the crystalline-metallic-ore-fat-soluble-compound complex is dried and stored for a period prior to being contacted with the wash solution.

Preferably the fat-soluble compound is present in the aqueous solution within a number of cells and the aqueous solution is a culture media.

Preferably the cells are those of *Dunaliella salina*.

- 20 Preferably the fat-soluble compound is a natural pigment.

Preferably the pigment is a carotenoid.

Preferably the carotenoid is beta-carotene.

- 4 -

Preferably the wash solution is an organic solvent.

Preferably the fat-soluble compound is isolated from the wash solution by evaporation or drying.

In another aspect of the present invention there is provided a substantially pure fat-soluble compound obtained using the method of any one of claims 1 to 13.

- 5 In yet another aspect of the present invention there is provided a crystalline-metallic-ore-fat-soluble-compound complex obtained using a method as herein described.

Figures

10 These and other aspects of the present invention, which should be considered in all its novel aspects, will become apparent from the following description of the preferred embodiment of the invention, which are given by way of example only, with reference to the accompanying figure in which:

Figure 1 illustrates a preferred extraction apparatus and method according to the present invention; and

15 Figure 2 illustrates a preferred extraction apparatus and alternative method according to the present invention.

Preferred Embodiment

20 The preferred embodiment of the invention is described below in terms of the recovery of beta-carotene from water containing the microalgal species *Dunaliella salina* (*D.salina*). It will be appreciated by those of general skill in the art that the invention would be applicable to the recovery of the other carotenoids and to other fat-soluble pigments from *D.salina*, and to the recovery of carotenoids or other fat-soluble compounds or pigments from other suitable organisms. The process of the present invention may also be applicable to the extraction from an aqueous solution of fat-soluble compounds suspended therein.

Throughout the following description the words adsorption and absorption, or derivatives thereof, such as adsorb or absorb, are used. The word adsorb is used to describe how a substance can be held on the surface of another and the word absorb to refer to the inclusion or incorporation of one substance into another. These words have been used interchangeably in the following text as the interactions between the substances (beta-carotene and magnetite) may be referred to in either way. In addition, the word attach is used to cover both adsorption and absorption. Those of general skill in the art will appreciate this factor.

General Principles of the Preferred Embodiment of the Invention

Beta-carotene, is one of a group of compounds called carotenoids; this group also includes alpha carotene, lutene, lutene monoepoxide, astaxanthin, zeaxanthin, canthaxanthin, and lycopene. These compounds are well characterised and those skilled in the art will recognise them as being coloured fat-soluble compounds which function as part of the light-capturing apparatus in photosynthetic pathways. Beta-carotene, in particular, is a precursor to vitamin A, a vitamin obtained from dietary sources, rather than *de novo*, in animals. In addition, the carotenoid family have been associated with antioxidant activities. As a result, carotenoids and in particular, beta-carotene, are sought after for use in many food and health products.

The microalgae *Dunaliella* are typically cultivated in water which has a high concentration of dissolved salts, particularly concentrates of seawater such as those used for the production of salt by solar evaporation. Such waters are very corrosive of metal which they come into contact with. Under conditions of optimal nutrient concentrations, moderate temperatures and intense solar radiation, *Dunaliella* can grow to concentrations of one million algal cells per ml. The individual cells can contain up to 10% of their weight as beta-carotene, and thus beta-carotene can accumulate to the extent of 15 mg per litre of brine. The rest of *Dunaliella* cell biomass is composed of protein, carbohydrates and other lipids.

The process of the present invention uses an absorption medium, magnetite, which absorbs beta-carotene with very high affinity, but does not absorb significant amounts of the other -- components of the cell mass. However, beta-carotene is a lipid which is contained within the cell membrane, as opposed to being secreted from the cell and thus free in solution, therefore

Dunaliella cell membranes must be disrupted before the beta-carotene is available for absorption.

Magnetite is a crystalline iron ore which has a surface of sharp edges with numerous cracks, crags and irregularities. It has been identified that in the present invention magnetite can be used for the dual purpose of disrupting cell membranes and absorbing beta-carotene. When *Dunaliella* cells are brought into contact with magnetite particles the cell membrane is punctured by the numerous sharp edges and the cell contents are disgorged into the bulk growth/culture medium.

Another hitherto undescribed property of magnetite is that, because of its unique structure, it selectively absorbs beta-carotene. This occurs as beta-carotene, being a lipid, is insoluble in water and the surface of magnetite crystals are somewhat hydrophobic in nature. When beta-carotene is present in an environment of brine and magnetite, it partitions towards the more hydrophobic solid phase than the hydrophilic liquid phase. When the surface of crags within the magnetite particle become coated in beta carotene, a more hydrophobic microenvironment is created in which further beta carotene is absorbed. The total loading of beta-carotene into magnetite is thus very high; for example 2% to 4% of the mass of the magnetite. At this concentration, the void spaces within the magnetite structures are filled with beta-carotene. It will be understood by those of general skill in the art that it will not always be appropriate to load the magnetite completely as it may impact on the downstream recovery of beta-carotene.

Yet another useful property of magnetite disclosed herein is that when beta-carotene is adsorbed onto, then absorbed into magnetite, the oxidation processes which ordinarily cause the rapid decay of beta-carotene, particularly when it is exposed to oxygen, are inhibited such that the magnetite/beta-carotene complex is very stable and does not degrade when exposed to heat or when it is dried.

Because of the above identified properties of magnetite which are not obvious, magnetite provides an ideal material upon which to collect and concentrate beta-carotene. However, it will be appreciated by those of general skill in the art that alternative absorption media, such as

another crystalline metallic ore which has properties equivalent to those of magnetite, for example hematite, may be used in the process of the present invention.

As indicated above when beta carotene has been absorbed on magnetite until the magnetite is saturated, the material is, for example, around 2% by weight beta-carotene. As the brine containing *D. salina* typically has a maximum beta-carotene concentration of 20 parts per million, absorption by the magnetite, in this example, thus concentrates the beta-carotene by a factor of one thousand fold. Magnetite typically has a bulk density of 4 Kg per litre whereas the brine used for growing beta-carotene-containing *D. salina* has a bulk density typically of 1.2 Kg per litre. Therefore by passing 1,000 litres of brine containing *D. salina* through one kilogram of magnetite, all the beta-carotene can be removed and contained in a volume of 250 ml, a concentration of almost 4,000 fold.

The beta-carotene can be easily desorbed from the magnetite by simply washing the magnetite with a suitable wash solution such as an organic solvent. Both polar and non-polar solvents are suitable for this purpose. Ordinarily non-polar solvents would not easily mix with a material such as magnetite when it is wetted with water or brine due to hydrophobicity. However, another useful feature of the microcrystalline structure of magnetite is that interfacial tension is broken by the sharp surface, thus a non-polar solvent is easily able to penetrate, and then dewater the magnetite.

The organic solvent used to desorb beta-carotene will contain essentially pure beta carotene, as other microalgal products are not absorbed onto the magnetite. Because the solvent contains pure beta-carotene it is particularly easy to remove the beta carotene and recover the solvent for re-use, for example by using reduced pressure devices such as crystallisers.

Solvents which are suitable for desorbing beta-carotene from magnetite include, but are not restricted to acetone, ethanol, hexane, petroleum ether, or any mixtures of these solvent.

Further, due to consumer demand for natural products it is preferable that natural solvents be used. In this regard we have found terpene alcohols to be efficient solvents for use in this invention; for example, cineol (eucalyptus oil), *d*-limonene (lemon oil), citral (citrus oil) and terpen-4-ol (tee tree oil).

Basic Apparatus and Extraction Example of the Preferred Embodiment of the Invention

An example of an apparatus in which the process of the preferred form of the invention may be conducted is shown in Diagram/Figure 1 and Figure 2. In Figure 1 the magnetite is contained within a conical vessel (101). The magnetite sits on a distribution plate, or plenum, (110) which separates the inlet pipe (103) from the vessel interior. This plenum (110) allows a solid phase of magnetite to settle onto the bottom of the vessel when the apparatus is not in use.

Brine containing *D. Salina* (102) is introduced at the bottom of the conical vessel via inlet pipe (103) at such a flow rate as to maintain the magnetite as a fluidised bed (104). Being a fluidised bed contactor, there is no prospect for the adsorption media, magnetite, to become clogged. The brine (containing cellular debris) (105) leaves the vessel at outlet pipe (106). It can either be sent into another similar vessel to (101) if it still contains unabsorbed Beta-carotene, or it can be returned to the algal growth pond.

One can see from Figure 1 that the fluidised bed separates into two different layers or phases. The bottom phase contains primarily magnetite particles and the top phase magnetite-beta-carotene complexes (109); which move upwards because of a change in their density due to forming the complex with beta-carotene. It will be appreciated by those of general skill in the art that the layers may not be distinct from one another as illustrated in Figures 1 and 2 and that while two layers do form they do so across a gradient as a result of the degree of loading of magnetite with beta-carotene. Further, it will be appreciated that the size of the magnetite particle and the velocity of fluid flowing into the vessel will have an effect on the position of that particle within the vessel. The Figures have been simplified to illustrate that magnetite loaded with beta-carotene will decrease in density during the process.

It should be noted that magnetite of various particle sizes may be used in the present invention. The size of such particles is not important in relation to the absorption of beta-carotene but it will have an effect on the behaviour of the fluidised bed (104). Thus, as a result of the particle size of the magnetite used the flow rate of solutions into the vessel (101) may be required to be altered to maintain the bed (104) in a fluidised state.

The beta-carotene can be desorbed from the magnetite in the top phase by changing the flow into the inlet at (103) from brine to the desorption solvent (107) of choice. A different flow rate is required to keep the magnetised bed fluidised due to the density differences between brine and the solvent. The solvent effluent (108) from (106) contains essentially pure beta-carotene which can be recovered by evaporating the solvent. During this stage magnetite present in the top phase may fall to the bottom phase as the beta-carotene is released and its density increases.

It can be clearly seen that by using a number of vessels such as (101), connected in series such that the outlet (106) of one is connected to the inlet (103) of the next vessel in the series, any number of vessels can be connected to each other. If both the inlet and the outlet are connected via a manifold which can feed either brine or desorption solvent into the vessel, then a continuous process cycle of adsorption/desorption/adsorption is possible.

Such a system operates at low pressure, has only valves as moving parts, can be constructed of cheap plastic material and has a very low energy requirement. As such, this system provides a very simple, efficient and reliable means of harvesting beta-carotene from brine.

Figure 2 uses the same apparatus as Figure 1 but illustrates an alternative embodiment in which when the magnetite has become progressively loaded with beta-carotene the magnetite-beta-carotene complex (109) is collected from the vessel (101) at outlet (106). At this stage the complex can be washed immediately with an appropriate solvent or stored at room temperature for prolonged periods without any significant deterioration of the contained beta-carotene and washed at a later date.

These basic examples will become further apparent from the specific examples 1 to 3 which follow.

Specific Examples relating to the Preferred Embodiment of the Invention

Example 1:

A culture of *Dunaliella salina* was grown in outdoor ponds containing sodium chloride at a concentration of 60g per litre (approximately 1 M). When the culture had attained a beta-carotene concentration of 9mg per litre, the culture was pumped into the bottom of a vertical

- 10 -

perspex cylinder of 100mm diameter at a rate of 1.5 litre per minute. When the cylinder became filled with liquid, 800g of magnetite (120 mesh) was introduced into the top of the cylinder.

The magnetic moved towards the bottom of the cylinder but became suspended within the cylinder as a fluidised bed which maintained a height of 400mm. When the fluidised bed

- 5 became stable, the culture which passed through the bed to the top of the cylinder was sampled and the beta carotene concentration was measured and found to be 3.9mg per litre.

While the culture medium was still being pumped through the bottom of the cylinder, a further 400g of magnetite was then introduced into the top of the cylinder. The fluidised bed then expanded to a height of 580mm. The culture emerging from the top of the cylinder was again

- 10 sampled and this time found to have a beta carotene concentration of 1.9mg per litre.

A further 400g of magnetite was then added to the cylinder which caused the fluidised bed height to increase to 750mm. At this bed height the culture emerging from the top of the bed appeared clear. The beta carotene concentration was measured and found to be 0.04mg per litre.

After approximately 1 hour of operation with a total added volume of magnetite of 1,600g, and a

- 15 constant upward culture medium flow rate of 1.5 litres per minute, the fluidised bed volume had expanded to 780mm, and had separated into two distinct zones. The upper zone had a slightly red colour and was 65mm high. The lower zone was the same black colour as the originally formed fluidised bed and was 715mm high. There was a distinct boundary between the two layers.

- 20 Magnetite material from the upper layer was collected using a pipette, then washed with fresh water and examined under a microscope. There was no sign of any algal cells adhering to this magnetite. The magnetite was then dried in a flow of warm air, weighed accurately and the washed with acetone. The acetone was collected and the beta carotene concentration in the acetone was determined by measuring the optical density at 450nm wavelength. It was
- 25 determined in this way that the magnetite contained 3.9% by weight beta carotene. ---

Example 2:

A culture of *Dunaliella salina* was grown in outdoor ponds containing sodium chloride at a

concentration of 60 g per litre (approximately 1 M) and magnesium chloride at 60g per litre (approximately 1 M). When the culture had attained a beta carotene concentration of 11mg per litre, the culture was pumped into the bottom of a vertical perspex cylinder of 100mm diameter at a rate of 1.4 litre per minute. When the cylinder became filled with liquid, 1,600g of magnetite (120 mesh) was introduced into the top of the cylinder. The magnetite moved towards the bottom of the cylinder but became suspended within the cylinder as a fluidised bed which maintained a height of 800 mm. When the fluidised bed became stable, the culture which passed through the bed to the top of the cylinder was sampled and the beta carotene concentration was measured and found to be 0.07mg per litre. The culture medium emerging from the top of the column was examined under a microscope. There were no intact algal cells observed, however cellular debris, comprising mostly broken cell membranes, and halobacteria were observed.

After approximately 2 hour of operation at a constant upward culture medium flow rate of 1.4 litres per minute, the fluidised bed volume had expanded to 845mm, and had separated into two distinct zones. The upper zone had a slightly red colour and was 165mm high. At this height the magnetic fluidised bed had reached the top of the perspex cylinder. As the bed expanded further, the top layer spilled over and was collected and was examined under a microscope. There was no sign of any algal cells adhering to this magnetite. The magnetite was then dried in a flow of warm air, weighed accurately and then washed with acetone. The acetone was collected and the beta carotene concentration in the acetone was determined by measuring the optical density at 450nm wavelength. In this way it was determined that the magnetite contained 3.8% by weight beta carotene.

Example 3:

A culture of *Dunaliella salina* was grown in outdoor ponds containing sodium chloride at a concentration of 90g per litre (approximately 1.5 M) and magnesium chloride at 90g per litre (approximately 1.5 M). When the culture had attained a beta carotene concentration of 14mg per litre, the culture was pumped into the bottom of a vertical perspex cylinder of 100mm diameter at a rate of 1.65 litre per minute. When the cylinder became filled with liquid, 1,600g of magnetite (120 mesh) was introduced into the top of the cylinder. The magnetic moved

towards the bottom of the cylinder but became suspended within the cylinder as a fluidised bed which maintained a height of 800mm. When the fluidised bed became stable, the culture which passed through the bed to the top of the cylinder was sampled and the beta carotene concentration was measured and found to be 0.06mg per litre.

- 5 In this example the cylinder was modified by creating a spillway 950mm up the length of the cylinder. After 95 minutes of operation, the upper (red) zone of the fluidised magnetite bed had reached the spillway, and magnetite began trickling from the spillway. This spilled magnetite was separated from the culture medium by decantation. The rate of flow of magnetite trickling from the cylinder was estimated by collecting the material for one minute, removing the culture medium by decantation and weighing the magnetite. It was found that approximately 600mg of magnetite was spilling from the cylinder each minute. By washing the magnetite with acetone and measuring the optical density of the washing acetone at 450nm, the spilled magnetite was found to contain 3.65% by weight beta carotene.

- 10 For the next 4 hours a 6g sample of fresh magnetite was added to the top of the cylinder every 10 minutes. The fresh magnetite could be seen to travel through the upper red zone into the lower black zone of the fluidised bed. For the 4 hours during which the trial was undertaken, the fluidised bed maintained a more-or-less constant height and a quite constant rate of red magnetite spillage from the spillway. At the completion of the trial, 400 litres of culture medium had been passed through the cylinder and substantially all the beta-carotene had been removed.

- 25 The fluid flow into the cylinder was then switched from culture medium at 1.65 litres per minute to cineole at a flow rate of 2.25 litres per minute. Again a fluidised bed was formed, this time with a bed height of 820mm. The cineole emerging from the top of the cylinder was a deep red colour. Spectrophotometric measurement of the cineole at 450nm showed it contained beta carotene at a concentration of 1.35% w/v. After about four minutes of flow, the cineole emerging from the top of the cylinder became a paler red, and after 6 minutes it was clear. All the eluted cineole was collected, and evaporated using a rotary evaporator. As the cineol evaporated, dark crystals of beta carotene were formed. When the cineole had completely

- 13 -

evaporated, the remaining crystalline material was collected and weighed. The material weighed 5.44g.

It can be concluded that the 400 litres of culture originally applied to the magnetite fluidised bed contained 5.6g of beta carotene. Of this, 5.44g was recovered in crystalline form from the
5 cineole eluent. This represents a recovery of over 97% of the original beta carotene present in the culture.

Industrial Application and Advantages

Carotenoids and other fat-soluble pigments are sought after additives for food and health products. The process of the present invention is very simple, requires little more energy than
10 that needed to reticulate water containing microalgae to the apparatus and thus provides an efficient means of extracting these compounds from their source and thus may prove of commercial and economic advantage.

The process has the further advantage of stabilising the product and enabling the convenient storage of the product as a concentrate.

FOOTNOTES

15

CLAIMS:

1. A method of extracting fat-soluble compounds from aqueous solutions including the steps:
providing an aqueous solution in which a fat-soluble compound is present;
providing a bed of crystalline metallic ore particles held in an appropriate vessel;
5 applying the aqueous solution to the bed of crystalline metallic ore particles substantially near
the bottom of the bed at a rate sufficient to form and maintain a fluidised bed of crystalline
metallic ore particles;
allowing the fat-soluble compound to attach to the crystalline metallic ore particles to form a
crystalline-metallic-ore-fat-soluble-compound complex;
10 providing a wash solution;
contacting the wash solution with the crystalline-metallic-ore-fat-soluble-compound complex
to desorb the fat-soluble compound from the complex;
collecting the wash solution containing the fat-soluble compound; and
isolating the fat-soluble compound from the wash solution.
- 15 2. A method as claimed in claim 1 wherein the crystalline metallic ore particles are magnetite
particles.
3. A method as claimed in any one of claims 1 or 2 wherein the fat-soluble compounds attach to
the metallic iron ore particles via means of adsorption or absorption.
- 20 4. A method as claimed in any one of claims 1 to 3 wherein the wash solution is contacted with
the crystalline-metallic-ore-fat-soluble-compound complex by applying the wash solution to
the fluidised bed of crystalline metallic ore particles substantially near the bottom of the
fluidised bed and at a rate sufficient to maintain the bed in a fluidised state and the resultant
wash solution containing the fat-soluble compound is collected from near the top of, or
above, the fluidised bed of crystalline metallic ore particles.
- 25 5. A method as claimed in any one of claims 1 to 3 further including the step of collecting the
crystalline-metallic-ore-fat-soluble-compound complex prior to providing a wash solution

- 15 -

and contacting the wash solution with the crystalline-metallic-ore-fat-soluble-compound complex.

6. A method as claimed in claim 5 wherein the crystalline-metallic-ore-fat-soluble-compound complex is collected from a region substantially near the top of the fluidised bed of crystalline metallic ore particles by means of continuous decantation.
7. A method as claimed in any one of claims 5 to 6 wherein the crystalline-metallic-ore-fat-soluble-compound complex is dried and stored for a period prior to being contacted with the wash solution.
8. A method as claimed in claim any one of claims 1 to 7 wherein the fat-soluble compound is present in the aqueous solution within a number of cells and the aqueous solution is a culture media.
9. A method as claimed in claim 8 wherein the cells are those of *Dunaliella salina*.
10. A method as claimed in any one of claims 1 to 9 wherein the fat-soluble compound is a natural pigment.
11. A method as claimed in claim 9 wherein the pigment is a carotenoid.
12. A method as claimed in claim 11 wherein the carotenoid is beta-carotene.
13. A substantially pure fat-soluble compound obtained using the method of any one of claims 1 to 12.
14. A crystalline-metallic-ore-fat-soluble-compound complex obtained using the method of claim 7.

FIG. 1

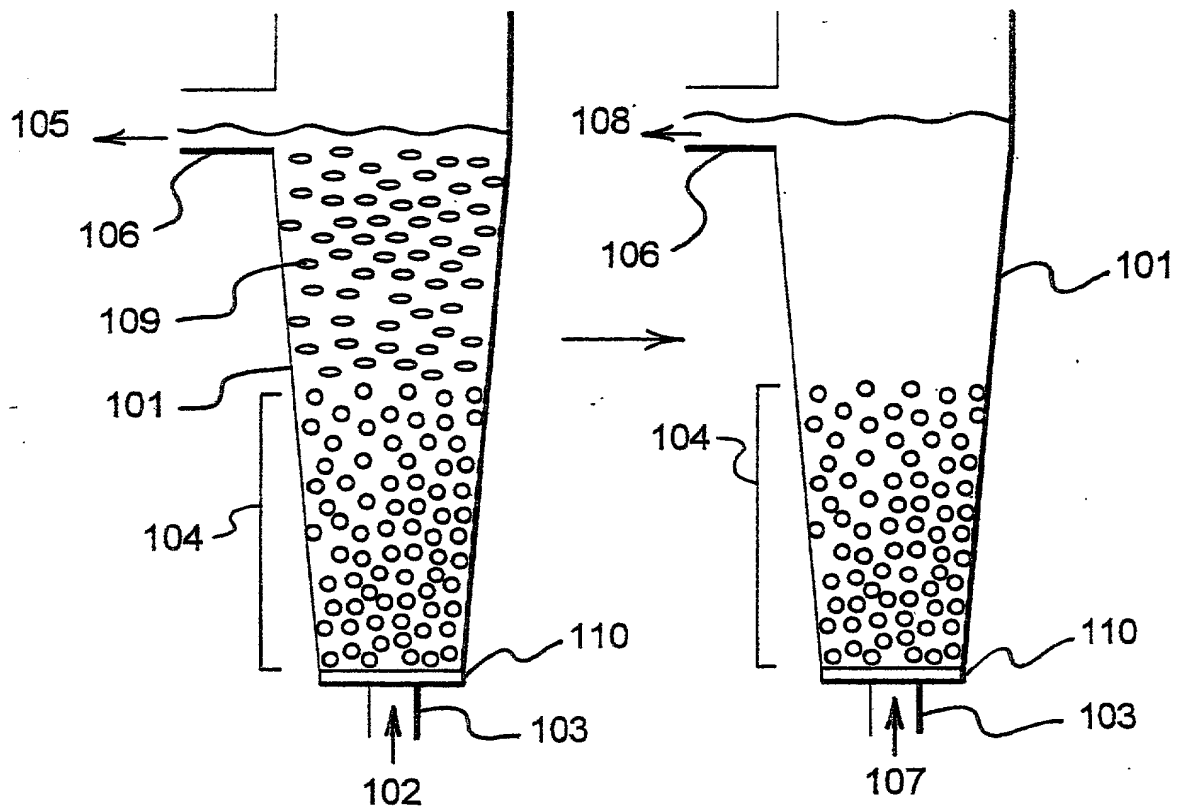
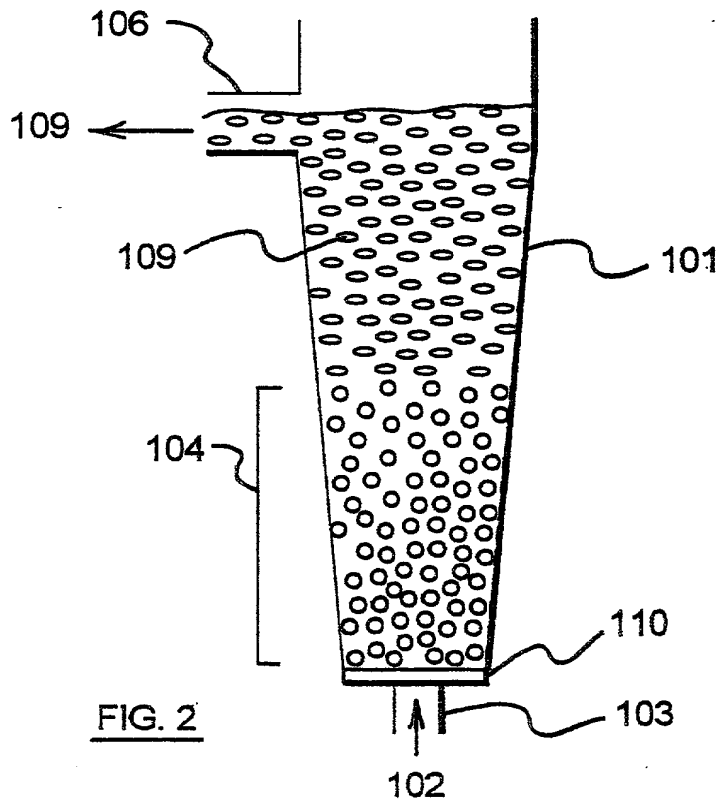


FIG. 2



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PIPERS (TAKAPUNA) NEW ZEALAND

No. 5232 P. 3/4

Combined Declaration and Power of Attorney

As a below named inventor, I hereby declare that :

My residence, post office address and citizenship are as stated below next to my name.

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled

A Method for Recovering Pigments from Algal Cultures

the specification of which

[X] was described and claimed in International Application # PCT/NZ00/00001, filed on 10 January 2000.

I hereby state that I have reviewed and understand the contents of the above identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information which is material to patentability as defined in Title 37, Code of Federal Regulations, §1.56.

I hereby claim foreign priority benefits under 35 USC 119 of any foreign application(s) for patent or inventor's certificate listed below and have also identified below any foreign application for patent or inventor's certificate having a filing date before that of the application on which priority is claimed.

Prior Foreign Application(s)

Priority Claimed

I hereby claim the benefit under 35 USC 120 of any United States application(s) listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States application in the manner provided by the first paragraph of 35 USC 112, I acknowledge the duty to disclose information which is material to patentability as defined in Title 37, Code of Federal Regulations, §1.56 which became available between the filing date of the prior application and the national or PCT international filing date of this application:

Application Serial # No	Filing Date	Status
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The undersigned hereby authorizes the US attorney or agent named herein to accept and follow instructions from **PIPER**s as to any action to be taken in the Patent and Trademark Office regarding this application without direct communication between the US attorney or agent and the undersigned. In the event of a change in the persons from whom instructions may be taken, the US attorney or agent named herein will be so notified by the undersigned.

As a named inventor, I hereby appoint the following attorneys to prosecute this application and transact all business in the Patent and Trademark Office connected therewith: **Robert J. PATCH, Reg. No. 17,355; Andrew J. PATCH, Reg. No. 32,925; Robert F. HARGEST, Reg. No. 25,590; Benoit CASTEL, Reg. No. 35,401, Eric JENSEN, Reg. No. 37,855 and Thomas W. PERKINS, Reg. No. 33,027 C/- YOUNG & THOMPSON, Second Floor, 745 South 23rd Street, Arlington, VA 22202.**

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